

CoraLite[®] 488-594 Phalloidin

Stock preparation

Prepare a 200T/mL stock solution by dissolving lyophilized phalloidin (300T) in 1.5mL sterile water and protect solution from light.

Fluorescence microscopy

1. Wash the cells 3 times with PBS.
2. Fix the cells on ice for 15 min with 4% paraformaldehyde in PBS.
Note: Avoid fixatives using methanol because they can destroy actin during the fixation process
3. Wash the cells 3 times with PBS
4. Permeabilize the cells with 0.2% Triton X-100 in PBS at room temperature for 5 min.
5. Wash the cells 3 times with PBS.
6. Prepare master mix by diluting 1-5 μ L working stock solution with 200 μ L PBS. For multiple samples, scale appropriately.
7. Add 200 μ L master mix to each sample, and incubate for 20 min at room temperature for staining. Protect from light.
Note: To avoid evaporation of the dye solution during the incubation, place samples in a humid environment.
8. Wash the cells 2-3 times with PBS.
9. Observe the samples under a fluorescence microscope
5. Wash the cells 3 times with PBS.