For Research Use Only

HA-Trap Agarose

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Catalog Number: ata

6 Publications

Basic Information

Catalog Number:
ata

Applications:
IP, Co-IP

Conjugate:
Agarose beads; ~90 um (cross-linked 4% agarose beads)

Host:
Alpaca

Type:
Nanobody

Class:
Recombinant

Description

The ChromoTek HA-Trap Agarose consists of an anti-HA tag Nanobody/VHH, which is coupled to agarose beads. It can be used for the immunoprecipitation of HA-tagged proteins from cell extracts of various organisms such as humans, mice, dogs, yeast, and plants.

Specificity/Target

Binds specifically to the HA-tag (sequence YPYDVPDYA) fused to a protein of interest at N-, C- or internal position. Please note that the affinity is highest for a C-terminal fusion. There is no cross-reactivity to other common peptide tags such as the His6-tag, FLAG-tag, Spot-Tag, V5-tag, Strep-tag, or C-tag (other tags not tested). Background binding to host cell proteins from a range of organisms such as human, mouse and dog cell lines or yeast and plants is low.

Elution buffer

2x SDS-sample buffer (Lammli)

Affinity (K_D)

6 nM for C-terminal HA-tags and ca. 180 nM for N-terminal fusions.

Storage

Storage:
Shipped at ambient temperature. Upon receipt store at +4°C. Stable for one year. Do not freeze!

Storage Buffer:
20% ethanol

For technical support and original validation data for this product please contact

T: 1 (888) 4PTGLAB (1-888-478-4522) (toll free in USA), or 1(312) 455-8498 (outside USA)

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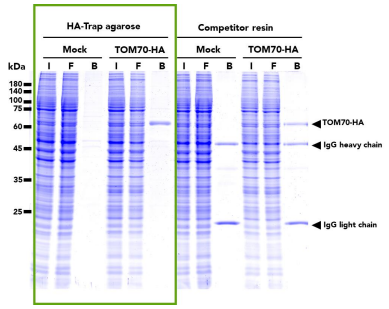
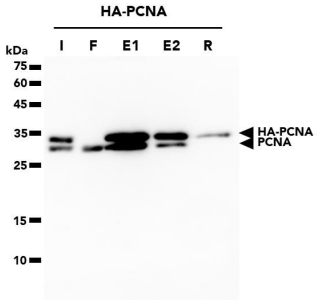
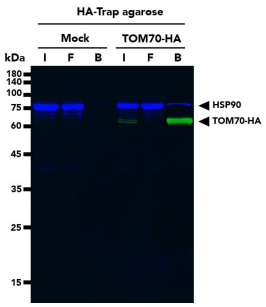
Selected Validation Data



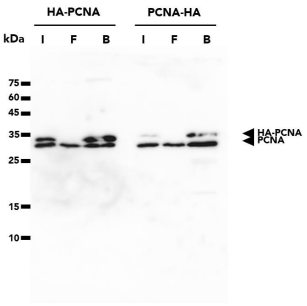
The HA-Trap Agarose (left) and a competitor resin (right) were used to immunoprecipitate TOM70-HA fusion protein from either untransfected (mock) HEK293T cells or HEK293T cell transfected with full-length TOM70-HA construct. Immunoprecipitation with HA-Trap Agarose results in clean, single-band pulldowns without any heavy and light chain contamination. SDS-PAGE analysis was done on samples from the Input (I), Flow-through (F), Bound (B) fra



The HA-Trap Agarose was used to immunoprecipitate HA-PCNA fusion protein from HEK293T cells. HA-PCNA protein was released from the trap through a two-step competitive elution with HA-peptide (ap). Samples from the Input (I), Flow-Through (F), 1st elution (E1), 2nd elution (E2), and residual (R) fractions were analyzed through WB. PCNA Monoclonal Antibody (60097-1-Ig) and Multi-rAb HRP-Goat Anti-Mouse Recombinant Secondary Antibody (RGAM001) were used in the WB analysis. Note: PCNA forms trimers resulting in co-elution of endogenous PCNA proteins with HA-tagged PCNA.



Co-IP using HA-Trap Agarose followed by multiplexed WB of TOM70-HA and HSP90 proteins from untransfected (mock) HEK293T cells and HEK293T cells transfected with full-length TOM70-HA construct. WB analysis was done on samples from the Input (I), Flow-through (F) and Bound (B) fractions of the IP. TOM70 Monoclonal Antibody (66593-1-Ig), Multi-rAB CoraLite Plus 488-Goat Anti-Mouse Recombinant Secondary Antibody (RGAM002), HSP90 Polyclonal Antibody (13171-1-AP), and Multi-rAb CoraLite Plus 750-Goat Anti Rabbit Recombinant Secondary Antibody (RGAR006) were used in the WB analysis.



The HA-Trap Agarose was used to immunoprecipitate HA-PCNA and PCNA-HA proteins from transfected HEK293T cells. WB analysis was done on samples from the Input (I), Flow-Through (F), and Bound (B) fractions of the IP using PCNA Monclonal Antibody (60097-1-Ig) and Multi-rAb HRP-Goat Anti-Mouse Recombinant Secondary Antibody (RGAM001). The HA-Trap is succesful in pulling down HA-tagged PCNA regardless of whether the tag is fused to the N- or C-terminal. Note: PCNA forms trimers, resulting in co-elution of endogenous PCNA with HA-tagged PCNA.