

## Speedy™ Human TNFSF18 One-Step ELISA Kit Datasheet

Please read it entirely before use

**Catalogue Number:** SE50206

**Size:** 96T

**Sensitivity:** 3.0 pg/mL

**Range:** 39-2500 pg/mL

**Usage:** For the quantitative detection of human TNFSF18 concentrations in serum and plasma.

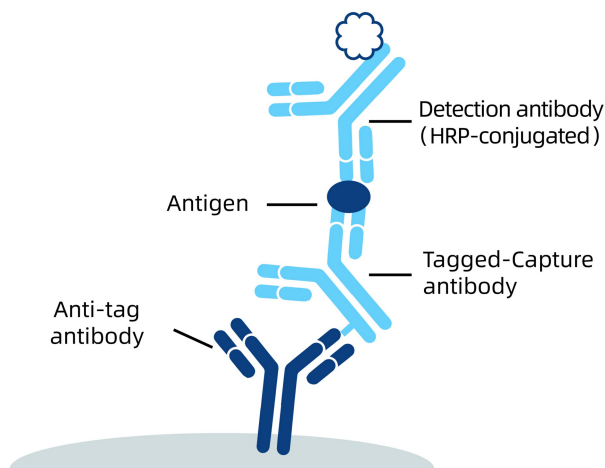
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## 1. Background

TNFSF18 is also named as Glucocorticoid-induced TNF-related ligand (hGITRL). GITR is a gene coding for a member of the TNF receptor superfamily. GITR activation by its ligand (GITRL) influences the activity of effector and regulatory T cells, thus participating in the development of immune response against tumours and infectious agents, as well as in autoimmune and inflammatory diseases. GITR-GITRL interactions co-stimulate both responder T-cell functions and the suppressive functions of TReg cells. GITRL is expressed at high levels in the small intestine, ovary, testis, kidney and endothelial cells.

## 2. Principle



An anti-tag antibody is pre-coated onto the bottom of wells. After adding antigen or samples, Tagged-Capture antibody and HRP-conjugated detection antibody, a sandwich complex is formed in the solution. TMB acts as a HRP substrate, and the solution color will change from colorless to blue. A stop solution containing sulfuric acid turns the solution yellow. The color intensity is proportional to the quantity of bound protein, which is measurable at 450 nm with the correction wavelength set at 630 nm.

## 3. Required Materials

- 3.1 A microplate reader capable of measuring absorbance at 450 nm with the correction wavelength set at 630 nm.
- 3.2 Calibrated, adjustable precision pipettes and disposable plastic tips. A manifold multi-channel pipette is recommended for large assays.
- 3.3 Plate washer: automated or manual.
- 3.4 Absorbent paper towels.
- 3.5 Glass or plastic tubes to prepare standard and sample dilutions.
- 3.6 Beakers and graduated cylinders.
- 3.7 Log-log or semi-log graph paper or computer and software for ELISA data analysis. A four-parameter logistic (4-PL) curve-fit is recommended. Proteintech data analysis website, <https://www.ptgcn.com/products/elisa-kits/>.
- 3.8 Microplate thermostatic shaker.

## 4. Kit Components and Storage

<b>Microplate</b> - 96 well microplate precoated an anti-tag antibody (8 well × 12 strips)	1 plate	<b>Unopened Kit:</b> Store at 2-8°C for 6 months or -20°C for 12 months. <b>Opened Kit:</b> All reagents stored at 2-8°C for 7 days. <b>Please use a new standard for each assay.</b>
<b>Protein standard</b> - 5000 pg/bottle; lyophilized	2 bottles	
<b>Capture antibody (100×)</b> - 60 µL/vial*	1 vial	
<b>Detection antibody, HRP-conjugated (100×)</b> - 60 µL/vial*	1 vial	
<b>Sample Diluent PT 1-ef</b> - 30 mL/bottle	1 bottle	
<b>Detection Diluent</b> - 15 mL/bottle	1 bottle	
<b>Wash Buffer Concentrate (20×)</b> - 30 mL/bottle	1 bottle	
<b>Tetramethylbenzidine Substrate (TMB)</b> - 12 mL/bottle	1 bottle	
<b>Stop Solution</b> - 12 mL/bottle	1 bottle	
<b>Plate Cover Seals</b>	4 pieces	

\* Centrifugation immediately before use

## 5. Safety Notes

- 5.1 Avoid any skin and eye contact with Stop Solution and TMB. In case of contact, wash thoroughly with water.
- 5.2 Do not use the kit after the expiration date.
- 5.3 Do not mix or substitute reagents or materials from other kit lots or other sources.
- 5.4 Be sure to wear protective equipment such as gloves, masks and goggles during the experiment.
- 5.5 When using an automated plate washer, adding a 30 second soak period following the addition of Wash Buffer to improve assay precision

## 6. Sample Collection and Storage

- 6.1 Serum: Allow blood samples to clot for 30 minutes, followed by centrifugation for 15 minutes at 1000×g. Clear serum can be assayed immediately or aliquoted and stored at -20°C. Avoid repeated freeze-thaw cycles.
- 6.2 Plasma: Use EDTA, heparin, or citrate as an anticoagulant for plasma collection. Centrifuge for 15 minutes at 1000×g within 30 minutes of collection. The plasma can be assayed immediately or aliquoted and stored at -20°C. Avoid repeated freeze-thaw cycles.

## 7. Regent Preparation

**7.1 Wash Buffer (1X):** If crystals have formed in the concentrate, warm to room temperature and mix gently until the crystals have completely dissolved. Add 30 mL of Wash Buffer Concentrate(20X) to 570 mL deionized or distilled water to prepare 1X Wash Buffer.

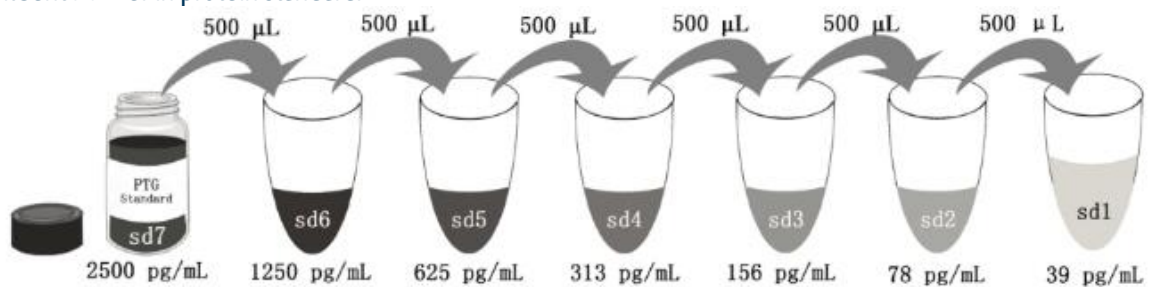
**7.2 Antibody Cocktail (1X):** Dilute 100X capture antibody and 100X HRP-conjugated detection antibody using Detection Diluent prior to assay. Suggested 1:100 dilution: 50  $\mu$ L 100X capture antibody + 50  $\mu$ L 100X Detection Antibody, HRP-conjugated + 4,900  $\mu$ L Detection Diluent. Mix gently but thoroughly.

**7.3 Sample Dilution:** Different samples should be diluted with corresponding Sample Diluent, samples may require further dilution if the readout values are higher than the highest standard OD reading. Variations in sample collection, processing and storage may affect the results of the measurement.

Recommended Dilution for different sample types: 1:2 or 1:4 is recommended for human serum and plasma.

### 7.4 Standard Serial Dilution:

Add 2 mL Sample Diluent PT 1-ef in protein standard.



Add # $\mu$ L of Standard diluted in the previous step	—	500 $\mu$ L	500 $\mu$ L	500 $\mu$ L	500 $\mu$ L	500 $\mu$ L	500 $\mu$ L
# $\mu$ L of Sample Diluent PT 1-ef	2000 $\mu$ L	500 $\mu$ L	500 $\mu$ L	500 $\mu$ L	500 $\mu$ L	500 $\mu$ L	500 $\mu$ L
	"sd7"	"sd6"	"sd5"	"sd4"	"sd3"	"sd2"	"sd1"

## 8. Assay Procedure Summary

Bring all reagents to room temperature before use (Detection antibody, HRP-conjugated antibody can be used immediately). To avoid cross-contamination, change pipette tips between additions of each standard level, between sample additions, and between reagent additions. Also, use separate reservoirs for each reagent.

8.1 Preset the layout of the microplate, including control group, standard group and sample group, take out the required number of microplate strips and return excess strips to the foil pouch containing the drying reagent pack and reseal; store at 4°C immediately. Microplate strips should be used in one week.

8.2 Add 50 µL standard or sample to appropriate wells. To avoid high background always add samples or standards to the well before the addition of antibody cocktail.

8.3 Add 50 µL 1× Antibody Cocktail solution (refer to Reagent Preparation 7.2) to each well. Seal plate with cover seal and incubate at 37°C on a microplate thermostatic shaker set at 400 rpm for 1 hour (incubate at 37°C for 2 hours is recommended if thermostatic shaker is not available) .

### 8.4 Wash

1) Gently remove the cover seal. Discard the liquid from wells by aspirating or decanting. Remove any residual solution by tapping the plate a few times on fresh paper towels.

2) Wash 4 times with 1× Wash Buffer, using at least 350-400 µL per well. Following the last wash, firmly tap plates on fresh towels 10 times to remove residual Wash Buffer. Avoid getting any towel fibers in the wells or wells drying out completely.

8.5 Add 100 µL TMB substrate solution to each well, protected from light. Incubate at 37°C on a microplate thermostatic shaker set at 400 rpm for 15 to 20 minutes. (Substrate Solution should remain colorless until added to the plate.)

8.6 Add 100 µL Stop Solution to each well in the same order as addition of the TMB substrate. Note: Avoid skin and eye contact with the Stop solution.

8.7 Read results immediately on a microplate reader at a wavelength of 450 nm. If possible, perform a double wavelength readout (450 nm and 630 nm).

8.8 Data analysis: Calculate the average of the duplicate readings (OD value) for each standard and sample, and subtract the average of the zero standard absorbance. Construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis, four-parameter logistic curve-fit (4-PL) analysis is recommended. If the samples have been diluted, the fitting result must be multiplied by the dilution factor used.

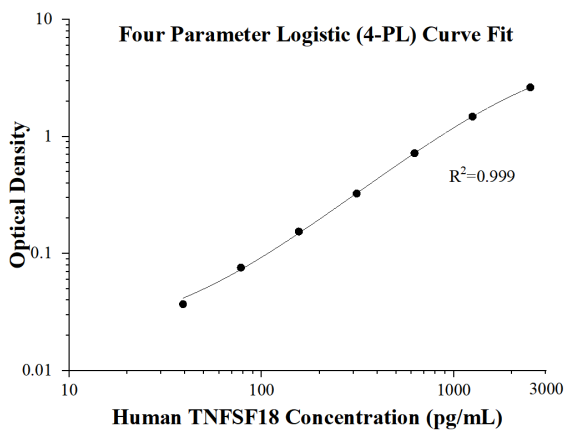
## Procedure summary



## 9. Validation Data

### 9.1 Standard curve

These standard curves are provided for demonstration only. A standard curve should be generated for each set of samples assayed.



(pg/mL)	O.D	Average	Corrected
0	0.05 0.0535	0.05175	-
39	0.091 0.0864	0.0887	0.03695
78	0.1281 0.1271	0.1276	0.07585
156	0.2009 0.2111	0.206	0.15425
313	0.3732 0.382	0.3776	0.32585
625	0.7756 0.7703	0.77295	0.7212
1250	1.5103 1.5615	1.5359	1.48415
2500	2.7191 2.6506	2.68485	2.6331

## 9.2 Precision

**Intra-assay Precision** (Precision within an assay) Three samples of known concentration were tested 8 times on one plate to assess intra-assay precision.

**Inter-assay Precision** (Precision between assays) Three samples of known concentration were tested in 16 separate assays to assess inter-assay precision.

Intra-assay Precision					Inter-assay Precision				
Sample	n	Mean (pg/mL)	SD	CV%	Sample	n	Mean (pg/mL)	SD	CV%
1	8	1,290.2	85.3	6.6	1	16	1,342.1	108.8	8.1
2	8	318.9	20.1	6.3	2	16	335.6	27.2	8.1
3	8	178.5	13.6	7.6	3	16	185.5	14.2	7.7

## 9.3 Recovery

The recovery of human TNFSF18 spiked to three different levels throughout the range of the assay was evaluated.

Sample Type		Average% of Expected	Range (%)
Human plasma	1:2	84	72-102

## 9.4 Sample values

**Human plasma** - Human plasma samples were evaluated for the presence of human TNFSF18 in this assay.

Sample Type	Mean (pg/mL)	%Detectable	Range (pg/mL)
Human plasma (n=16)	192.4	31.3	ND-243.7

ND\*=Non-detectable

## 9.5 Sensitivity

The minimum detectable dose of human TNFSF18 is 3.0 pg/mL. This was determined by adding two standard deviations to the concentration corresponding to the mean O.D. of 20 zero standard replicates.

## 9.6 Linearity

To assess the linearity of the assay, human plasma samples were spiked with high concentrations of human TNFSF18 in various matrices and diluted with the appropriate **Sample Diluent** to produce samples with values within the dynamic range of the assay.

		Human plasma
1:2	Average% of Expected	74
	Range (%)	71-77
1:4	Average% of Expected	97
	Range (%)	94-100
1:8	Average% of Expected	94
	Range (%)	91-98
1:16	Average% of Expected	91
	Range (%)	87-96

## 9.7 Specificity

This assay recognizes natural and recombinant human TNFSF18.

The following factors prepared at 50 ng/mL were assayed and exhibited no cross-reactivity or interference.

Recombinant human:

APRIL

LIGHT

TNF- $\alpha$

TNF- $\beta$

TRAIL

Recombinant mouse:

GITR

## 10. References

1. Giuseppe Nocentini. et al. GITR/GITRL: more than an effector T cell co-stimulatory system. (2007) Eur J Immunol. 37(5):1165-9.
2. Ethan M Shevach. et al. The GITR-GITRL interaction: co-stimulation or contrasuppression of regulatory activity. (2006) Nat Rev Immunol. 6(8):613-8.
3. Pedro M L. et al. Glucocorticoid-induced tumor necrosis factor receptor family-related ligand triggering upregulates vascular cell adhesion molecule-1 and intercellular adhesion molecule-1 and promotes leukocyte adhesion. (2013) J Pharmacol Exp Ther. 347(1):164-72.