

Speedy™ Mouse Adiponectin One-Step ELISA Kit Datasheet

Please read it entirely before use

Catalogue Number: SE60003

Size: 96T

Sensitivity: 1.4 pg/mL

Range: 15.6-1000 pg/mL

Usage: For the quantitative detection of mouse Adiponectinin concentrations in serum, plasma, cell culture supernatant and tissue homogenate.

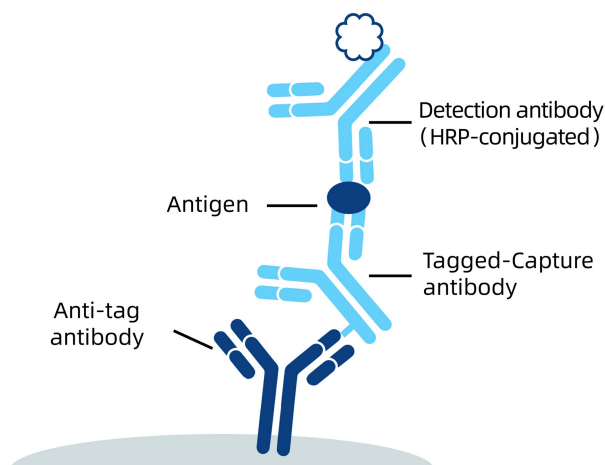
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1. Background

Adiponectin, also known as Acrp30, AdipoQ, and GBP28, is an adipocyte-secreted protein that circulates in high concentrations in the serum and acts to increase insulin sensitivity. Adiponectin modulates a number of metabolic processes, including glucose regulation and fatty acid oxidation. The serum concentration of adiponectin decreases in obese patients, opposite to other adipocytokines. In recent studies, serum adiponectin level progressively decreased in patients according to the severity of obesity, insulin resistance, diabetes mellitus and cardiovascular diseases. Low serum adiponectin level may be a predictive factor for type 2 diabetes mellitus (T2DM) and other cardiovascular diseases.

2. Principle



An anti-tag antibody is pre-coated onto the bottom of wells. After adding antigen or samples, Tagged-Capture antibody and HRP-conjugated detection antibody, a sandwich complex is formed in the solution. TMB acts as a HRP substrate, and the solution color will change from colorless to blue. A stop solution containing sulfuric acid turns the solution yellow. The color intensity is proportional to the quantity of bound protein, which is measurable at 450 nm with the correction wavelength set at 630 nm.

3. Required Materials

- 3.1 A microplate reader capable of measuring absorbance at 450 nm with the correction wavelength set at 630 nm.
- 3.2 Calibrated, adjustable precision pipettes and disposable plastic tips. A manifold multi-channel pipette is recommended for large assays.
- 3.3 Plate washer: automated or manual.
- 3.4 Absorbent paper towels.
- 3.5 Glass or plastic tubes to prepare standard and sample dilutions.
- 3.6 Beakers and graduated cylinders.
- 3.7 Log-log or semi-log graph paper or computer and software for ELISA data analysis. A four-parameter logistic (4-PL) curve-fit is recommended. Proteintech data analysis website, <https://www.ptgcn.com/products/elisa-kits/>.
- 3.8 Microplate thermostatic shaker.

4. Kit Components and Storage

Microplate - 96 well microplate precoated an anti-tag antibody (8 well × 12 strips)	1 plate	Unopened Kit: Store at 2-8°C for 6 months or -20°C for 12 months. Opened Kit: All reagents stored at 2-8°C for 7 days. Please use a new standard for each assay.
Protein standard - 2000 pg/bottle; lyophilized	2 bottles	
Capture antibody (100×) - 60 µL/vial*	1 vial	
Detection antibody, HRP-conjugated (100×) - 60 µL/vial*	1 vial	
Sample Diluent PT 4B1 - 30 mL/bottle	2 bottles	
Detection Diluent - 15 mL/bottle	1 bottle	
Wash Buffer Concentrate (20×) - 30 mL/bottle	1 bottle	
Tetramethylbenzidine Substrate (TMB) - 12 mL/bottle	1 bottle	
Stop Solution - 12 mL/bottle	1 bottle	
Plate Cover Seals	4 pieces	

* Centrifugation immediately before use

5. Safety Notes

- 5.1 Avoid any skin and eye contact with Stop Solution and TMB. In case of contact, wash thoroughly with water.
- 5.2 Do not use the kit after the expiration date.
- 5.3 Do not mix or substitute reagents or materials from other kit lots or other sources.
- 5.4 Be sure to wear protective equipment such as gloves, masks and goggles during the experiment.
- 5.5 When using an automated plate washer, adding a 30 second soak period following the addition of Wash Buffer to improve assay precision

6. Sample Collection and Storage

- 6.1 Serum: Allow blood samples to clot for 30 minutes, followed by centrifugation for 15 minutes at 1000×g. Clear serum can be assayed immediately or aliquoted and stored at -20°C. Avoid repeated freeze-thaw cycles.
- 6.2 Plasma: Use EDTA, heparin, or citrate as an anticoagulant for plasma collection. Centrifuge for 15 minutes at 1000×g within 30 minutes of collection. The plasma can be assayed immediately or aliquoted and stored at -20°C. Avoid repeated freeze-thaw cycles.
- 6.3 Cell Culture Supernatant: Remove particulates by centrifugation for 5 minutes at 500×g and assay immediately or aliquot and store samples at ≤ -20°C. Avoid repeated freeze-thaw cycles.
- 6.4 Tissue Homogenates: The preparation of tissue homogenates will vary depending upon tissue type. Rinse tissue with 1 × PBS to remove excess blood, Then add 1 mL of 1 × PBS per 100 mg tissue, homogenized and stored overnight at -20°C. After two freeze-thaw cycles were performed to break the cell membranes, the homogenates were centrifuged for 5 minutes at 10000 × g at 4°C. Collect the supernatant, assay immediately or aliquot and store at -20°C. Avoid repeated freeze-thaw cycles.

7. Regent Preparation

7.1 Wash Buffer (1X): If crystals have formed in the concentrate, warm to room temperature and mix gently until the crystals have completely dissolved. Add 30 mL of Wash Buffer Concentrate(20X) to 570 mL deionized or distilled water to prepare 1X Wash Buffer.

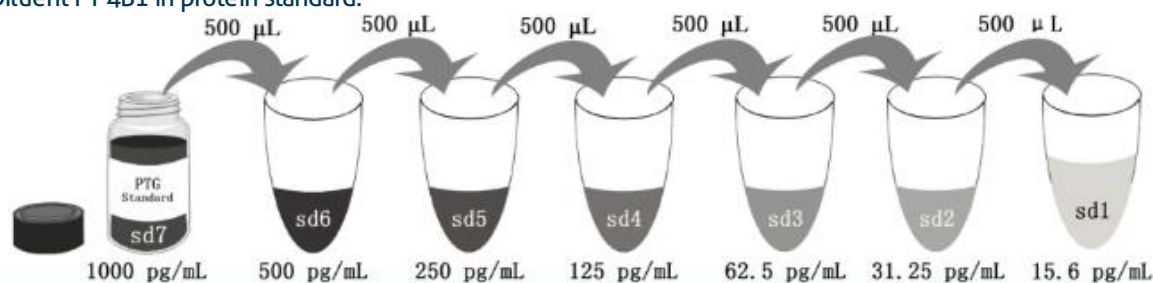
7.2 Antibody Cocktail (1X): Dilute 100X capture antibody and 100X HRP-conjugated detection antibody using Detection Diluent prior to assay. Suggested 1:100 dilution: 50 μ L 100X capture antibody + 50 μ L 100X Detection Antibody, HRP-conjugated + 4,900 μ L Detection Diluent. Mix gently but thoroughly.

7.3 Sample Dilution: Different samples should be diluted with corresponding Sample Diluent, samples may require further dilution if the readout values are higher than the highest standard OD reading. Variations in sample collection, processing and storage may affect the results of the measurement.

Recommended Dilution for different sample types: 1:40,000 or 1:80,000 is recommended for mouse serum and plasma. 1:2 or 1:4 is recommended for cell culture supernatant. 1:200 or 1:400 is recommended for tissue homogenate.

7.4 Standard Serial Dilution:

Add 2 mL Sample Diluent PT 4B1 in protein standard.



Add # μ L of Standard diluted in the previous step	—	500 μ L	500 μ L	500 μ L	500 μ L	500 μ L	500 μ L
# μ L of Sample Diluent PT 4B1	2000 μ L	500 μ L	500 μ L	500 μ L	500 μ L	500 μ L	500 μ L
	"sd7"	"sd6"	"sd5"	"sd4"	"sd3"	"sd2"	"sd1"

8. Assay Procedure Summary

Bring all reagents to room temperature before use (Detection antibody, HRP-conjugated antibody can be used immediately). To avoid cross-contamination, change pipette tips between additions of each standard level, between sample additions, and between reagent additions. Also, use separate reservoirs for each reagent.

8.1 Preset the layout of the microplate, including control group, standard group and sample group, take out the required number of microplate strips and return excess strips to the foil pouch containing the drying reagent pack and reseal; store at 4°C immediately. Microplate strips should be used in one week.

8.2 Add 50 µL standard or sample to appropriate wells. To avoid high background always add samples or standards to the well before the addition of antibody cocktail.

8.3 Add 50 µL 1X Antibody Cocktail solution (refer to Reagent Preparation 7.2) to each well. Seal plate with cover seal and incubate at 37°C on a microplate thermostatic shaker set at 400 rpm for 1 hour (incubate at 37°C for 2 hours is recommended if thermostatic shaker is not available) .

8.4 Wash

1) Gently remove the cover seal. Discard the liquid from wells by aspirating or decanting. Remove any residual solution by tapping the plate a few times on fresh paper towels.

2) Wash 4 times with 1X Wash Buffer, using at least 350-400 µL per well. Following the last wash, firmly tap plates on fresh towels 10 times to remove residual Wash Buffer. Avoid getting any towel fibers in the wells or wells drying out completely.

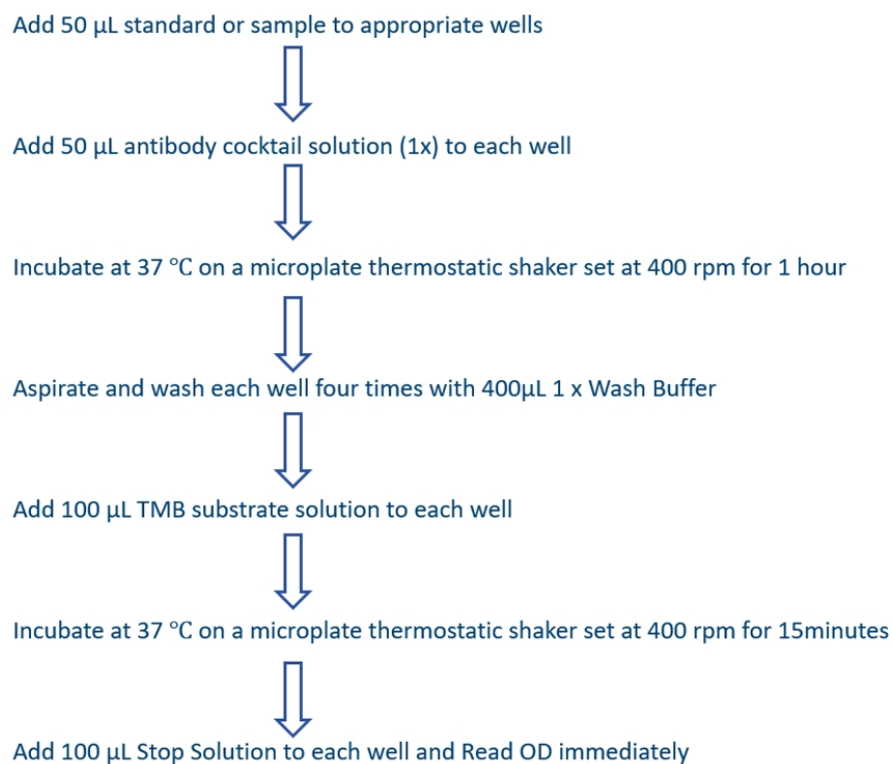
8.5 Add 100 µL TMB substrate solution to each well, protected from light. Incubate at 37°C on a microplate thermostatic shaker set at 400 rpm for 15 to 20 minutes. (Substrate Solution should remain colorless until added to the plate.)

8.6 Add 100 µL Stop Solution to each well in the same order as addition of the TMB substrate . **Note: Avoid skin and eye contact with the Stop solution.**

8.7 Read results immediately on a microplate reader at a wavelength of 450 nm. If possible, perform a double wavelength readout (450 nm and 630 nm).

8.8 Data analysis: Calculate the average of the duplicate readings (OD value) for each standard and sample, and subtract the average of the zero standard absorbance. Construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis, four-parameter logistic curve-fit (4-PL) analysis is recommended. If the samples have been diluted, the fitting result must be multiplied by the dilution factor used.

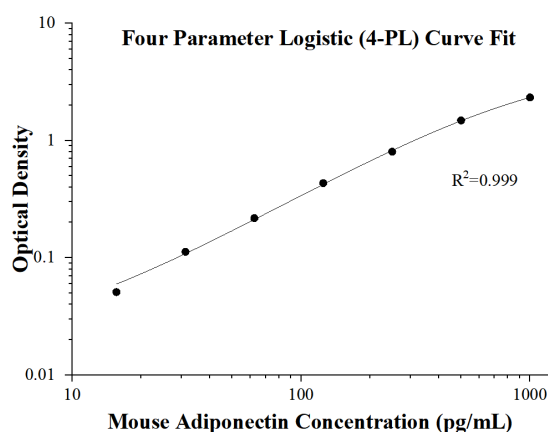
Procedure summary



9. Validation Data

9.1 Standard curve

These standard curves are provided for demonstration only. A standard curve should be generated for each set of samples assayed.



(pg/mL)	O.D	Average	Corrected
0	0.0735 0.077	0.07525	-
15.6	0.1223 0.1301	0.1262	0.05095
31.25	0.1856 0.1899	0.18775	0.1125
62.5	0.2906 0.295	0.2928	0.21755
125	0.5094 0.507	0.5082	0.43295
250	0.8765 0.88	0.87825	0.803
500	1.5594 1.5611	1.56025	1.485
1000	2.4155 2.3946	2.40505	2.3298

9.2 Precision

Intra-assay Precision (Precision within an assay) Three samples of known concentration were tested 8 times on one plate to assess intra-assay precision.

Inter-assay Precision (Precision between assays) Three samples of known concentration were tested in 16 separate assays to assess inter-assay precision.

Intra-assay Precision					Inter-assay Precision				
Sample	n	Mean (pg/mL)	SD	CV%	Sample	n	Mean (pg/mL)	SD	CV%
1	8	477.3	14.4	3.0	1	16	495.4	24.9	5.0
2	8	125.0	4.5	3.6	2	16	127.8	6.3	4.9
3	8	63.0	2.1	3.3	3	16	64.1	3.1	4.8

9.3 Recovery

The recovery of mouse Adiponectin spiked to three different levels throughout the range of the assay was evaluated.

Sample Type		Average% of Expected	Range (%)
Mouse serum	1:100,000	103	97-108
	1:200,000	99	96-104
Cell culture supernatant	1:8	113	103-123
	1:16	102	96-112
Tissue homogenate	1:400	93	84-101
	1:800	90	87-92

9.4 Sample values

Mouse serum - Mouse serum samples were evaluated for the presence of mouse Adiponectin in this assay.

Sample Type	Mean (µg/mL)	Range (µg/mL)
Mouse serum (n=16)	14.1	5.7-24.6

Cell culture supernatant - Mouse lung (1 lung, 1-2 mm pieces) were cultured for 3 days in RPMI1640 plus 15% fetal bovine serum. An aliquot of cell culture supernatants was removed, assayed for mouse Adiponectin and measured 693.2 pg/mL.

Tissue homogenate - Mouse liver were rinsed with PBS to remove excess blood, chopped into 1-2 mm pieces, homogenized in 5-10 mL of PBS in a tissue homogenizer, and stored at ≤ -80°C 5 min. After two freeze-thaw cycles were performed to break the cell membranes, the homogenates were centrifuged for 5 minutes at 5000 x g to remove particulate. Homogenates from liver tissue were assayed for mouse Adiponectin and measured 46.4 ng/mL.

9.5 Sensitivity

The minimum detectable dose of mouse Adiponectin is 1.4 pg/mL. This was determined by adding two standard deviations to the concentration corresponding to the mean O.D. of 20 zero standard replicates.

9.6 Linearity

To assess the linearity of the assay, samples were diluted with the appropriate **Sample Diluent** to produce samples with values within the dynamic range of the assay.

(The mouse serum was initially diluted 1:20,000. The tissue homogenate was initially diluted 1:100.)

		Mouse serum	Cell culture supernatant	Tissue homogenate
1:2	Average% of Expected	100	100	100
	Range (%)	-	-	-
1:4	Average% of Expected	101	108	102
	Range (%)	99-103	103-112	102-103
1:8	Average% of Expected	101	116	104
	Range (%)	100-102	108-124	103-105
1:16	Average% of Expected	105	122	105
	Range (%)	103-108	118-126	99-111

9.7 Specificity

This assay recognizes natural and recombinant mouse Adiponectin.

The following factors prepared at 50 ng/mL were assayed and exhibited no cross-reactivity or interference.

Recombinant mouse:

TNF- α

CD30 Ligand

Fas Ligand

OX40 Ligand

A sample containing 50 ng/mL of the recombinant mouse CD40 Ligand reads as 21.83 pg/mL (0.04% cross-reactivity).

10. References

1. Wang ZV. et al. (2016) J Mol Cell Biol. 8(2):93-100.
2. Liu M. et al. (2014) Best Pract Res Clin Endocrinol Metab.28(1):25-31.
3. Díez JJ. et al. (2003) Eur J Endocrinol.148(3):293-300.
4. Yamauchi T. et al. (2003) Nature.423(6941):762-9.