

Mouse ICAM-1/CD54 Sandwich ELISA Kit Datasheet

Please read it entirely before use

Catalogue Number: KE10099

Size: 5*96T

Sensitivity: 0.02 ng/mL

Range: 0.156-10 ng/mL

Usage: For the quantitative detection of mouse ICAM-1/CD54 concentrations in serum, plasma, cell culture supernatant and tissue homogenate.

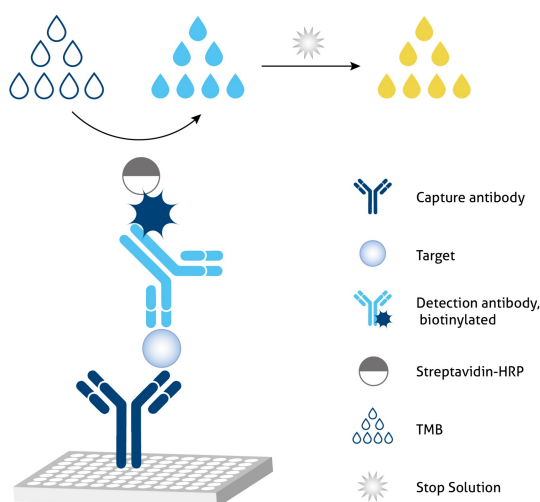
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1. Background

Intercellular adhesion molecule 1 (ICAM-1, also known as CD54) is a transmembrane glycoprotein of the immunoglobulin superfamily and is critical for the firm attachment and transmigration of leukocytes out of blood vessels and into tissues. ICAM-1 is expressed by several cell types, typically on endothelial cells and cells of the immune system, and its expression can be up-regulated by various stimuli, including TNF- α , INF- γ , IL-1 and thrombin. It is a ligand for LFA-1 and Mac-1, serves as a receptor for rhinovirus, and is one of several receptors used by *Plasmodium falciparum*. ICAM-1 can exist as membrane-bound form (mICAM-1) and soluble form (sICAM-1). The sICAM-1 arises from alternative splicing and proteolysis of mICAM-1 and appears to be associated with leukocyte activation to produce LFA-1.

2. Principle



Sandwich ELISA structure (Detection antibody labeled with biotin)

A capture antibody is pre-coated onto the bottom of wells which binds to analyte of interest. A detection antibody labeled with biotin also binds to the analyte. Streptavidin-HRP binds to the biotin. TMB acts as the HRP substrate and the solution color will change from colorless to blue. A stop solution containing sulfuric acid turns solution yellow. The color intensity is proportional to the quantity of bound protein which is measurable at 450 nm with the correction wavelength set at 630 nm.

3. Required Materials

- 3.1 A microplate reader capable of measuring absorbance at 450 nm with the correction wavelength set at 630 nm.
- 3.2 Calibrated, adjustable precision pipettes and disposable plastic tips. A manifold multi-channel pipette is recommended for large assays.
- 3.3 Plate washer: automated or manual.
- 3.4 Absorbent paper towels.
- 3.5 Glass or plastic tubes to prepare standard and sample dilutions.
- 3.6 Beakers and graduated cylinders.
- 3.7 Log-log or semi-log graph paper or computer and software for ELISA data analysis. A four-parameter logistic (4-PL) curve-fit is recommended.

4. Kit Components and Storage

Microplate - antibody coated 96-well microplate (8 well × 12 strips)	5 plates	Unopened Kit: Store at 2-8°C for 6 months or -20°C for 12 months. Opened Kit: All reagents stored at 2-8°C for 7 days. Please use a new standard for each assay.
Protein standard - 20 ng/bottle; lyophilized	10 bottles	
Detection Antibody (100×), biotinylated - 600 µL/vial*	1 vial	
Streptavidin-horseradish peroxidase (HRP) (100×) - 600 µL/vial*	1 vial	
Sample Diluent PT 4B1 - 150 mL/bottle	2 bottles	
Detection Diluent - 150 mL/bottle	1 bottle	
Wash Buffer Concentrate (20×) - 150 mL/bottle	1 bottle	
Tetramethylbenzidine Substrate (TMB) - 60 mL/bottle	1 bottle	
Stop Solution - 60 mL/bottle	1 bottle	
Plate Cover Seals	15 pieces	

* Centrifugation immediately before use

5. Safety Notes

- 5.1 Avoid any skin and eye contact with Stop Solution and TMB. In case of contact, wash thoroughly with water.
- 5.2 Do not use the kit after the expiration date.
- 5.3 Do not mix or substitute reagents or materials from other kit lots or other sources.
- 5.4 Be sure to wear protective equipment such as gloves, masks and goggles during the experiment.
- 5.5 When using an automated plate washer, adding a 30 second soak period following the addition of Wash Buffer to improve assay precision

6. Sample Collection and Storage

- 6.1 Serum: Allow blood samples to clot for 30 minutes, followed by centrifugation for 15 minutes at 1000xg. Clear serum can be assayed immediately or aliquoted and stored at -20°C. Avoid repeated freeze-thaw cycles.
- 6.2 Plasma: Use EDTA, heparin, or citrate as an anticoagulant for plasma collection. Centrifuge for 15 minutes at 1000xg within 30 minutes of collection. The plasma can be assayed immediately or aliquoted and stored at -20°C. Avoid repeated freeze-thaw cycles.
- 6.3 Cell Culture Supernatant: Remove particulates by centrifugation for 5 minutes at 500xg and assay immediately or aliquot and store samples at ≤ -20°C. Avoid repeated freeze-thaw cycles.
- 6.4 Tissue Homogenates: The preparation of tissue homogenates will vary depending upon tissue type. Rinse tissue with 1 x PBS to remove excess blood, Then add 1 mL of 1 x PBS per 100 mg tissue, homogenized and stored overnight at -20°C. After two freeze-thaw cycles were performed to break the cell membranes, the homogenates were centrifuged for 5 minutes at 10000 x g at 4°C. Collect the supernatant, assay immediately or aliquot and store at -20°C. Avoid repeated freeze-thaw cycles.

7. Regent Preparation

7.1 Wash Buffer (1X): If crystals have formed in the concentrate, warm to room temperature and mix gently until the crystals have completely dissolved. Add 30 mL of Wash Buffer Concentrate(20X) to 570 mL deionized or distilled water to prepare 1X Wash Buffer.

7.2 Detection Antibody(1X): Dilute 100X Detection Antibody 1:100 using Detection Diluent prior to assay. Suggested 1:100 dilution: 10 μ L 100X Detection Antibody + 990 μ L Detection Diluent (Centrifuge the 100 X Detection Antibody solution for a few seconds prior to use).

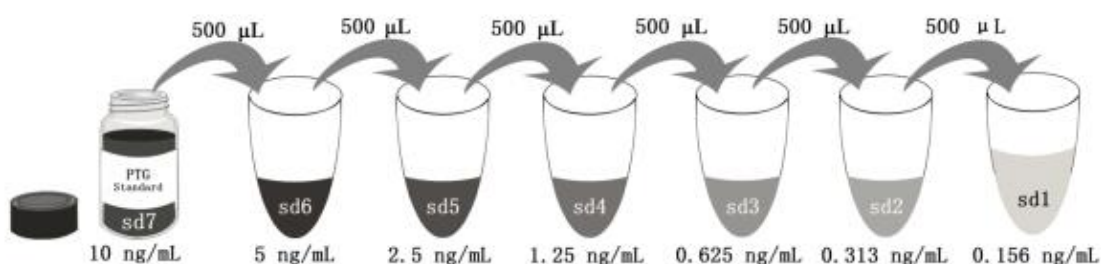
7.3 Streptavidin-HRP(1X): Dilute 100X Streptavidin-HRP 1:100 using Detection Diluent prior to assay. Suggested 1:100 dilution: 10 μ L 100X Streptavidin-HRP + 990 μ L Detection Diluent (Centrifuge the 100X Streptavidin-HRP solution for a few seconds prior to use).

7.4 Sample Dilution: Different samples should be diluted with corresponding Sample Diluent, samples may require further dilution if the readout values are higher than the highest standard OD reading. Variations in sample collection, processing and storage may affect the results of the measurement.

Recommended Dilution for different sample types: 1:320 or 1:640 is recommended for mouse serum and plasma; 1:2 or 1:4 is recommended for cell culture supernatant; 1:160 to 1:4,000 is recommended for tissue homogenates.

7.5 Standard Serial Dilution:

Add 2 mL Sample Diluent PT 4B1 in protein standard.



Add # μ L of Standard diluted in the previous step	—	500 μ L	500 μ L	500 μ L	500 μ L	500 μ L	500 μ L
# μ L of Sample Diluent PT 4B1	2000 μ L	500 μ L	500 μ L	500 μ L	500 μ L	500 μ L	500 μ L
	"sd7"	"sd6"	"sd5"	"sd4"	"sd3"	"sd2"	"sd1"

8. Assay Procedure Summary

Bring all reagents to room temperature before use (Detection antibody and Streptavidin-HRP can be used immediately). To avoid cross-contamination, change pipette tips between additions of each standard level, between sample additions, and between reagent additions. Also, use separate reservoirs for each reagent.

8.1 Take out the required number of microplate strips and return excess strips to the foil pouch containing the drying reagent pack and reseal; store at 4°C immediately. Microplate strips should be used in one week.

8.2 Preset the layout of the microplate, including control group, standard group and sample group, add 100 μ L of each standard and sample to the appropriate wells. (Make sure sample addition is uninterrupted and completed within 5 to 10 minutes, It is recommended to assay all standards, controls, and samples in duplicate).

8.3 Seal plate with cover seal, pressing it firmly onto top of microwells. Incubate the plate for 2 hours at 37°C.

8.4 Wash

1) Gently remove the cover seal. Discard the liquid from wells by aspirating or decanting. Remove any residual solution by tapping the plate a few times on fresh paper towels.

2) Wash 4 times with 1X Wash Buffer, using at least 350-400 μ L per well. Following the last wash, firmly tap plates on fresh towels 10 times to remove residual Wash Buffer. Avoid getting any towel fibers in the wells or wells drying out completely.

8.5 Add 100 μ L of 1X Detection Antibody solution (refer to Reagent Preparation 7.2) to each well. Seal plate with cover seal and incubate for 1 hour at 37°C.

8.6 Repeat wash step in 8.4.

8.7 Add 100 μ L of 1X Streptavidin-HRP solution (refer to Reagent Preparation 7.3) to each well. Seal plate with cover seal and incubate the plate for 40 minutes at 37°C.

8.8 Repeat wash step in 8.4.

8.9 Signal development: Add 100 μ L of TMB substrate solution to each well, protected from light. Incubate for 15 to 20 minutes. Substrate Solution should remain colorless until added to the plate.

8.10 Quenching color development: Add 100 μ L of Stop Solution to each well in the same order as addition of the TMB substrate. Mix by tapping the side of the plate gently. NB: Avoid skin and eye contact with the Stop solution.

8.11 Read results: Immediately after adding Stop solution read the absorbance on a microplate reader at a wavelength of 450 nm. If possible, perform a double wavelength readout (450 nm and 630 nm).

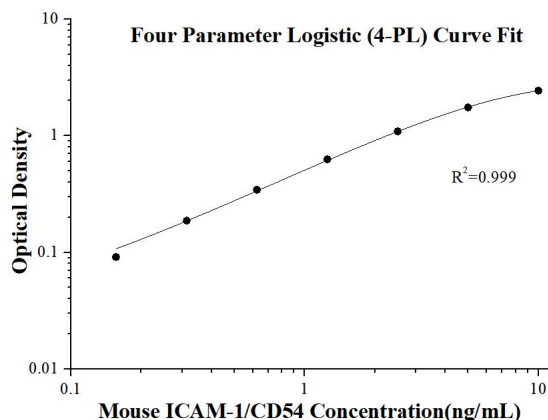
8.12 Data analysis: Calculate the average of the duplicate readings (OD value) for each standard and sample, and subtract the average of the zero standard absorbance. Construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis, use four-parameter logistic curve-fit (4-PL) analysis to do this. If the samples have been diluted, the OD readout from the standard curve must be multiplied by the dilution factor used.

Step	Reagent	Volume	Incubation	Wash	Notes
1	Standard and Samples	100 μ L	120 min	4 times	Cover Wells incubate at 37°C
2	Diluent Antibody Solution	100 μ L	60 min	4 times	Cover Wells incubate at 37°C
3	Diluent HRP Solution	100 μ L	40 min	4 times	Cover Wells incubate at 37°C
4	TMB Substrate	100 μ L	15-20 min	Do not wash	Incubate in the dark at 37°C
5	Stop Solution	100 μ L	0 min	Do not wash	-
6	Read plate at 450 nm and 630 nm immediately after adding Stop solution. DO NOT exceed 5 minutes.				

9. Validation Data

9.1 Standard curve

These standard curves are provided for demonstration only. A standard curve should be generated for each set of samples assayed.



(ng/mL)	O.D	Average	Corrected
0	0.039 0.038	0.039	-
0.156	0.132 0.128	0.130	0.091
0.313	0.223 0.229	0.226	0.187
0.625	0.383 0.382	0.383	0.344
1.25	0.678 0.657	0.668	0.629
2.5	1.139 1.120	1.130	1.091
5	1.822 1.757	1.789	1.751
10	2.523 2.437	2.480	2.441

9.2 Precision

Intra-assay Precision (Precision within an assay) Three samples of known concentration were tested 20 times on one plate to assess intra-assay precision.

Inter-assay Precision (Precision between assays) Three samples of known concentration were tested in 24 separate assays to assess inter-assay precision.

Intra-assay Precision				
Sample	n	Mean (ng/mL)	SD	CV%
2	20	1.10	0.02	1.74
3	20	0.27	0.01	3.90
1	20	4.80	0.15	3.09

Inter-assay Precision				
Sample	n	Mean (ng/mL)	SD	CV%
1	24	4.72	0.17	3.61
2	24	1.14	0.04	3.21
3	24	0.30	0.02	5.72

9.3 Recovery

The recovery of mouse ICAM-1/CD54 spiked to three different levels throughout the range of the assay in various matrices was evaluated.

Sample Type		Average% of Expected	Range (%)
Mouse serum	1:1,000	85	80-89
	1:2,000	87	80-91
Cell culture supernatant	1:2	91	82-95
	1:4	94	83-98
Tissue homogenate	1:500	97	88-115
	1:1,000	95	89-101

9.4 Sample values

Mouse serum -mouse serum samples were evaluated for the presence of mouse ICAM-1/CD54 in this assay.

Sample Type	Mean (ng/mL)	Range (ng/mL)
Mouse serum (n=16)	544.56	467.24-655.89

Cell culture supernatant:

Two mouse lungs (1-2 mm pieces in 40 mL of medium) were cultured for 6 days in RPMI supplemented with 10% fetal bovine serum. An aliquot of conditioned media was removed, assayed for mouse ICAM-1/CD54, and measured 2.50 ng/mL.

J774A.1 mouse reticulum cell sarcoma macrophage cells (1×10^6 cells/mL) were cultured for 1 day in DMEM containing 10% fetal bovine serum and stimulated with 100 ng/mL of recombinant mouse IFN- γ and 1.0 μ g/mL LPS. An aliquot of the cell culture supernate was removed, assayed for mouse ICAM-1/CD54, and measured 6.12 ng/mL.

Tissue homogenate - Organs from 2 mice were rinsed with PBS to remove excess blood, chopped into 1-2 mm pieces, homogenized in 5-10 mL of PBS in a tissue homogenizer, and stored at $\leq -80^\circ\text{C}$ 5 min. After two freeze-thaw cycles were performed to break the cell membranes, the homogenates were centrifuged for 5 minutes at 5000 x g to remove particulate. Homogenates from spleen and liver were assayed for mouse ICAM-1/CD54 and measured 209.31 ng/mL and 6,120.37 ng/mL, respectively.

9.5 Sensitivity

The minimum detectable dose of mouse ICAM-1/CD54 is 0.02 ng/mL. This was determined by adding two standard deviations to the concentration corresponding to the mean O.D. of 20 zero standard replicates.

9.6 Linearity

To assess the linearity of the assay, samples were diluted with the appropriate **Sample Diluent** to produce samples with values within the dynamic range of the assay.

(The mouse serum samples were initially diluted 1:160. The tissue homogenate was initially diluted 1:80.)

		Mouse serum	Cell culture supernatant	Tissue Homogenate
1:2	Average% of Expected	100	100	100
	Range (%)	-	-	-
1:4	Average% of Expected	110	104	108
	Range (%)	105-115	101-107	106-109
1:8	Average% of Expected	115	112	115
	Range (%)	111-117	109-116	114-116
1:16	Average% of Expected	119	116	102
	Range (%)	113-126	111-124	93-114

10. References

1. Lawson C, et al. (2009) Pharmacol Rep. 61(1):22-32.
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