

Flow Cytometry Cytoplasmic Staining Protocol

Reagents required:

4% PFA Fix Buffer (4% paraformaldehyde dissolved in 1x PBS, adjust pH to 7.4)

Flow Cytometry Perm Buffer (10x) (PF00011-C) dilute to 1x concentrate in distilled water prior to use

Flow Cytometry Staining Buffer (1x) (PF00012)

1x PBS

Flow cytometry antibodies

Experiment procedures:

- 1. Harvest cells and wash them twice with 1x PBS by centrifugation at $350-500 \times g$ for 5 minutes each time, discard the supernatant.
- 2. (Optional) Perform cell surface staining with recommended amount of fluorochrome-conjugated primary antibody, wash the cells with 1 mL staining buffer by centrifugation at $350-500 \times g$ for 5 minutes, discard the supernatant.
- 3. Resuspend the cells with 200 μ L 4% PFA fix buffer and vortex briefly, incubate for 20 minutes at room temperature in the dark.
- 4. Centrifuge at 350-500 x g for 5 minutes, discard the supernatant.
- 5. Resuspend the cells with 2 mL of 1x Flow Cytometry Perm Buffer and incubate at room temperature for 5 minutes in the dark.
- 6. Centrifuge at 350-500 x g for 5 minutes, discard the supernatant.
- 7. Resuspend the cells with 100 µL of 1x Flow Cytometry Perm Buffer.
- 8. Add the recommended amount of primary antibody for detection of intracellular target and incubate for 20-60 minutes at 4°C in the dark.
- 9. Wash the cells with 1 mL of 1x Flow Cytometry Staining Buffer by centrifugation at 350-500 x g for 5 minutes, discard the supernatant.

Note: If using fluorochrome-conjugated primary antibodies, skip to step 12.

- 10. Resuspend the cells with diluted fluorochrome-conjugated secondary antibody in 100 μ L of 1x Flow Cytometry Perm Buffer and incubate for 15-30 minutes at 4°C in the dark.
- 11. Wash the cells with 1 mL of 1x Flow Cytometry Staining Buffer by centrifugation at 350-500 x g for 5 minutes, discard the supernatant.
- 12. Resuspend the cells with 200-500 µL of 1x Flow Cytometry Staining Buffer and analyze on flow cytometer.