

Phantom Dyes Viability Staining Protocol

Buffers: 1X PBS (azide-free, protein/serum free) | Stain Buffer - 1X PBS with 2% FBS, 0.09% Sodium Azide

1. Remove Phantom Dye vial from freezer and thaw at room temperature.
2. Spin Phantom Dye vial down to collect contents at bottom before opening.
3. Prepare Stain Buffer for labeling procedure.
4. Wash cells twice in 1-2 mL 1X PBS. Spin at 300-400 x g for 5 minutes at room temperature. Afterwards, decant supernatant.
5. Resuspend to 1-10 x 10⁶ cells /mL in 1X PBS.
6. Add 1 μ L of Phantom Dye solution for each 1 mL of cell suspension and vortex immediately.
7. Incubate for 30 minutes at 2-8°C in the dark to prevent photobleaching.
8. Wash cells 1-2 times with 1-2 mL Stain Buffer to remove unreacted dye.
9. Cells can be subsequently stained, fixed and permeabilized according to user protocol.

Note: Cells labeled with Phantom Dyes can be cryopreserved for later use or used in intracellular staining protocols without any loss of fluorescence intensity.